



BHS

Belgian Hematology Society

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Transfusion: Practical Aspects & Hemovigilance

BHS

02/03/2024

Pr H. El Kenz



Plan

- **Transfusion of blood components**
 - **Transfusion of packed red blood cells (PRBC)**
 - Description of component
 - Pretransfusion Compatibility testing
 - **Transfusion of fresh frozen plasma (FFP)**
 - Description of component
 - Pretransfusion Compatibility testing
 - **Transfusion of platelets**
 - Description of component
 - Pretransfusion Compatibility testing

➤ **Hemovigilance**

Transfusion of blood components

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➤ Hemovigilance

Transfusion of packed red blood cells (PRBC)

Description of component

- Centrifugation of whole blood (removal of plasma and buffy-coat) or red cell apheresis
- Primary citrated anticoagulant (red cells shelf life \pm 21 days)
- Additive solutions (extension of shelf life to 42-49 days)
- PRBC volume is depending on the method of preparation
- PRBC (Council of Europe)
 - HCT of 0,50-0,70
 - Hb of minimum 40g/unit
 - Haemolysis at the end of the storage $<0,8\%$ of red cell mass
 - Leucocyte count $< 1 \times 10^6$ per unit
- Conservation at $+2^{\circ}\text{C}$ to $+6^{\circ}\text{C}$
- Administration through a 170-200 μm filter



Transfusion of packed red blood cells (PRBC)

Description of component

- PRBC
- PRBC Irradiated
- PRBC CMV negative

- PRBC cryopreserved (-60°C to -80°C)
- PRBC washed
- PRBC for neonates or baby packs (O-; CMV-)
- PRBC for intra-uterine transfusion (O-; CMV-; irradiated; <7days; HCT: 0,75-0,85)



Transfusion of packed red blood cells (PRBC)

Description of component

- **Irradiation of PRBC:**
 - TA-GVHD (Transfusion-associated graft-versus-host disease) is a very rare complication following transfusion of viable allogeneic lymphocytes into immunosuppressed recipients
 - Irradiation of PRBC for the prevention of TA-GVHD by inactivating residual lymphocytes with gamma rays or X-rays at a minimum dose of 25 Gy

bjh guidelines

Guidelines on the use of irradiated blood components

Theodora Foukaneli,^{1,2} Paul Kerr,³ Paula H.B. Bolton-Maggs,^{4,5} Rebecca Cardigan,⁶ Alasdair Coles,⁷ Andrew Genney,⁸ David Jane,⁹ Dinakantha Kumararatne,¹⁰ Ania Manson,¹⁰ Helen V. New,^{11,12} Nicholas Torpey¹³ and on behalf of the British Society for Haematology Guidelines Transfusion Task Force

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BSH updated UK guidelines 2020

bjh guideline

Guideline development for prevention of transfusion-associated graft-versus-host disease: reduction of indications for irradiated blood components after prestorage leukodepletion of blood components

Johanna C. Wiersum-Osselton,¹ Jennichjen Slomp,² J. H. Frederik Falkenburg,³ Tessa Geltink,⁴ Hans L. P. van Duijnhoven,⁵ Tanja Netelenbos⁶ and Martin R. Schipperus⁷

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Dutch updated guidelines 2021

Transfusion of PRBC

Description of component



Table I. Indications for irradiation: key differences between the BSH and Dutch guidelines.

BSH updated UK guidelines 2020 ²	Dutch updated guidelines 2021 ¹
Hodgkin lymphoma at any stage of the disease should receive irradiated components indefinitely	Hodgkin lymphoma stage III or IV (with bone marrow infiltration) not routinely required (depends on medications)
All patients treated with purine analogue drugs should receive irradiated components indefinitely	Use of purine/pyrimidine agonists Patients with long-lasting T-cell suppression after medication: fludarabine or other T-cell-depleting medication if the approved product information warns of TA-GVHD risk, for six months after cessation of the therapy
Stem cell transplants Allografts: to receive irradiated components from start of conditioning and be continued until defined criteria are met ² rather than a fixed time.	Stem cell transplants (time increased from earlier guidelines) Allogeneic stem cell transplantation: until one year after last medication/intervention
Autografts for three months (six months if total body irradiation was part of conditioning)	Autologous stem cell transplantation: until six months after transplantation
Irradiation required from seven days before and during bone marrow/stem cell harvest	Peripheral blood stem cell apheresis and bone marrow collection: no longer required
Patients with aplastic anaemia undergoing treatment with antithymocyte globulin or alemtuzumab (anti-CD52) should receive irradiated components duration not defined	Medication which in combination with patient's illness gives a long-lasting T-cell suppression, such as anti-CD52 treatments for haematological diseases and antithymocyte treatment for aplastic anaemia: from the initiation of treatment till six months after completing treatment
Routine irradiation of red cells to preterm or term infants not required unless there was a previous intrauterine transfusion	Premature babies and/or pregnancy <32 weeks (previously up to six months after due date) No longer required
Both: Intra-uterine transfusions (IUT), thereafter until six months after the due date	

bjh commentary

Guidelines: the same evidence but different conclusions — relaxation of indications for irradiation of cellular blood components?

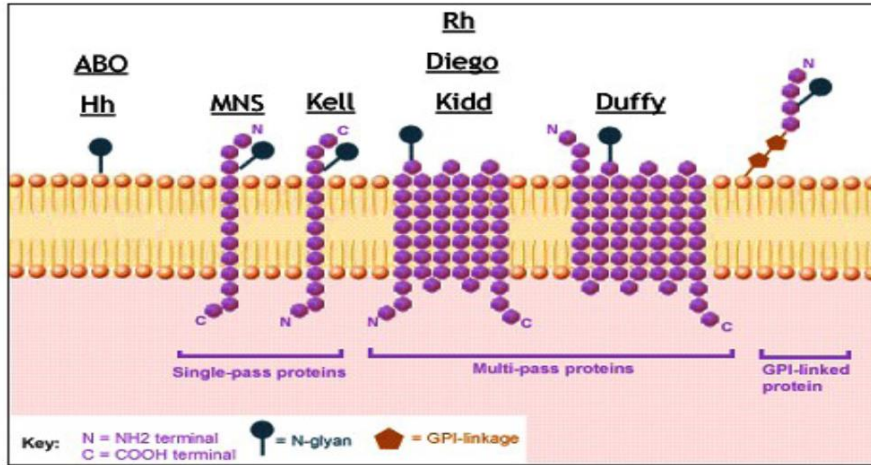
Paula H. B. Bolton-Maggs

Faculty of Biology, Medicine and Health, University of Manchester, Manchester, UK

Commentary on Wiersum-Oostbeek et al. Guideline development for prevention of transfusion-associated graft-versus-host disease: reduction of indications for irradiated blood components after prestorage leukodepletion of blood components. *Brit J Haematol.* 2021;195:681–688.

Transfusion of packed red blood cells (PRBC)

Pretransfusion Compatibility testing



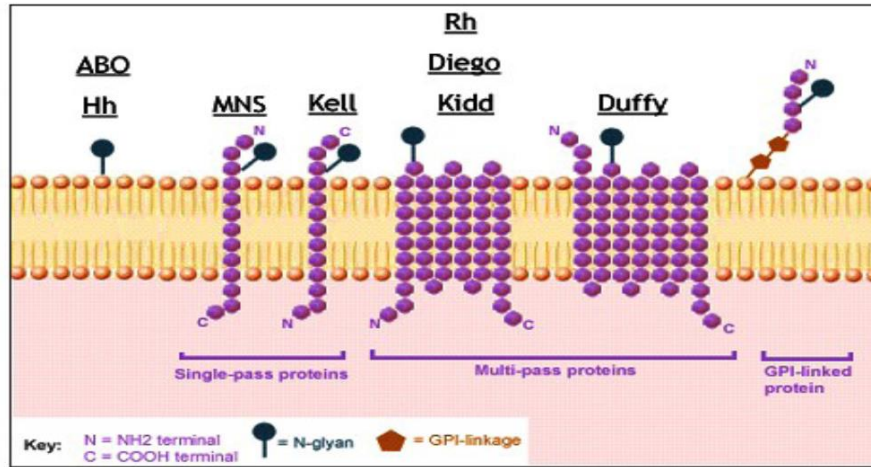
Currently recognized antigens classification:

- Systems
- Collections
- Low incidence antigens
- High incidence antigens

<http://www.isbtweb.org/working-parties/red-cell-immunogenetics-and-blood-group-terminology/>

Transfusion of packed red blood cells (PRBC)

Pretransfusion Compatibility testing



Patient antibodies:

- Natural antibodies
- Immune antibodies
- Auto-antibodies

Transfusion of packed red blood cells (PRBC)

Pretransfusion Compatibility testing

- Pre-transfusion tests:
 - Major Crossmatch
 - Patient's ABO/Rh types (2x) and crossreaction of serum's patient with ABO compatible PRBC
 - If negative: PRBC are reserved to the patient
 - If positive: red cell antibody identification and major crossmatch; information of the physician in case of short blood supply
 - Type and Screen
 - Patient's ABO/Rh types (2x) and Indirect Antiglobulin Test (IAT)
 - If positive IAT: Antibody identification and Crossmatch of antigen-negative PRBC; information of the physician in case of short blood supply
 - If negative IAT: labelling of the ABO compatible PRBC at the moment of the blood order

Transfusion of packed red blood cells (PRBC)

Pretransfusion Compatibility testing



TRANSFUSION MEDICINE Official Journal of the British Blood Transfusion Society

Transfusion Medicine | GUIDELINES

Guidelines for pre-transfusion compatibility procedures in blood transfusion laboratories*

British Committee for Standards in Haematology
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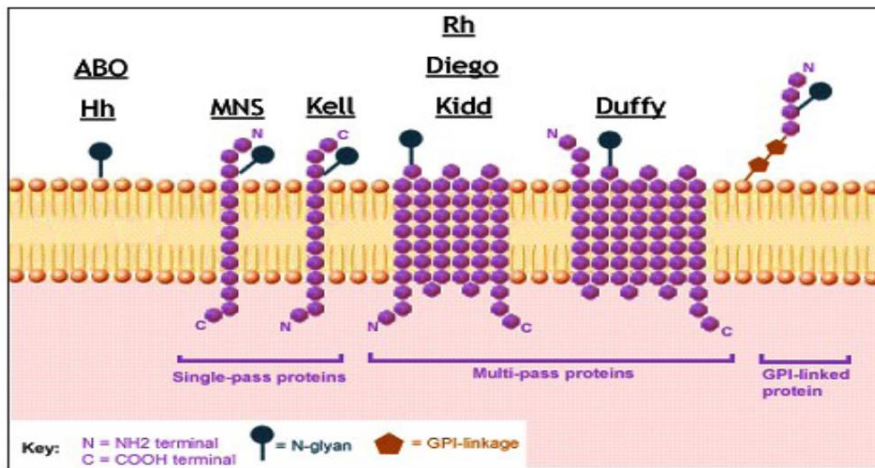
Received 18 July 2012; accepted for publication 27 September 2012

Key Recommendation 12:

Unless secure electronic patient identification systems are in place, a second sample should be requested for confirmation of the ABO group of a first time patient prior to transfusion, where this not impede the delivery of urgent red cells or other components

Transfusion of packed red blood cells (PRBC)

Pretransfusion Compatibility testing



Alloimmunization through transfusion or pregnancy

Incidence of alloimmunization in the general population: 2% to 5%

Incidence alloimmunization in patients with SCD: 30% to 40%

Administration of PRBC to SCD patients

- ❑ Alloimmunization to red blood cell remains a serious complication
- ❑ Risk factors associated with alloimmunization:
 - ❑ **Antigenic differences between donors and SCD patients**
 - ❑ HLA II genotype influence predisposition to the RBC antibody responder status
 - ❑ Inflammatory state of SCD patients

Administration of PRBC to SCD patients

Antigenic differences between donors and SCD patients

- Polymorphic differences in immunogenic RBC antigens between caucasian donors and african transfusion recipients
 - I. Significantly different RBC antigen frequencies of some common blood groups (RH, KEL, FY, JK, MNS,...)
 - II. RH variants
 - III. Loss of High-incidence antigens → Rare blood groups
 - IV. Low-incidence antigens → Alloimmunization in intra-ethnic transfusions

Administration of PRBC to SCD patients

Antigenic differences between donors and SCD patients

➤ Polymorphic differences in immunogenic RBC antigens between caucasian donors and african transfusion recipients

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Administration of PRBC to SCD patients

Antigenic differences between donors and SCD patients

	Caucasian frequency %	African frequency %	BRU+HUDE %
Fya -	34	90	92
Fy(a -b -)	Very rare	68	92
Jkb -	26	50	67
S -	45	70	67
Kell 6 ou Jsa	< 1	20	NT

Phenotype C-,E-,K-, Fy^{a-}, Jk^{b-}: 26% of Africans; <2% of Caucasians

Administration of PRBC to SCD patients

Antigenic differences between donors and SCD patients

➤ Polymorphic differences in immunogenic RBC antigens between caucasian donors and african transfusion recipients

I. Significantly different RBC antigen frequencies of some common blood groups (RH, KEL, FY, JK, MNS,...)

II. RH variants

III. Loss of High-incidence antigens → Rare blood groups

IV. Low-incidence antigens → Alloimmunization in intra-ethnic transfusions

Administration of PRBC to SCD patients

Antigenic differences between donors and SCD patients

- Loss of a High-incidence antigens:
 - RH:-18, RH:-34, RH:-46 → 0,1% in Africans
 - KEL:-7 → 1% in Africans
 - MNS:-5 → 1% in Africans
 -

Short blood supply in Europe

Administration of PRBC to SCD patients

Antigenic differences between donors and SCD patients

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Administration of PRBC to SCD patients

HLA II genotype influence predisposition to the RBC antibody responder status

- ❑ Some patients develop alloantibodies following initial RBC transfusions while others do not even after multiple transfusions
- ❑ RBC antibody responder and RBC antibody non-responder

HLA type and risk of alloimmunization in sickle cell disease

Carolyn Hoppe,^{1*} William Klitz,^{2,3} Elliott Vichinsky,¹ and Lori Styles¹

Red blood cell (RBC) transfusions are frequently required to treat patients with sickle cell disease (SCD) [1]. One of the most serious complications of repeat transfusion is alloimmunization to RBC antigens [2]. Because human leukocyte antigen (HLA) genes mediate the response to foreign antigens, particular HLA alleles may predispose to the development of alloimmunization in patients with SCD who receive multiple transfusions. We conducted a case-control study to determine if particular HLA alleles are associated with alloimmunization and whether HLA homozygosity influences the risk of developing RBC alloantibodies. High-resolution HLA genotyping was performed on DNA samples from 159 multiply transfused patients with SCD. HLA allele frequencies were compared between alloantibody-positive and alloantibody-negative groups. The *HLA-DRB1*1503* allele was associated with an increased risk ($P = 0.039$), while *HLA-DRB1*0901* conferred protection from alloimmunization ($P = 0.008$). HLA Class II locus homozygosity was more frequently observed in the alloantibody-negative group ($P = 0.01$). These preliminary findings suggest that particular *HLA-DRB1* alleles and overall homozygosity at HLA class II loci are associated with alloimmunization risk in SCD. If confirmed, HLA type may serve as a useful genetic predictor of alloimmunization risk, and permit a targeted approach to the use of phenotypically matched blood.

American Journal of Hematology

Administration of PRBC to SCD patients


- ❑ Alloimmunization to red blood cell remains a serious complication
- ❑ Risk factors associated with alloimmunization:
 - ❑ Antigenic differences between donors and SCD patients
 - ❑ HLA II genotype influence predisposition to the RBC antibody responder status
 - ❑ **Inflammatory state of SCD patients**

Administration of PRBC to SCD patients

- ❑ SCD is characterized by chronic inflammation
- ❑ Hypothesis: “Inflammation may play a role in the high rate of alloimmunization”

bjh research paper

Prevalence and risk factors for red blood cell alloimmunization in 175 children with sickle cell disease in a French university hospital reference centre

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Denise Amiranoff,⁵ Jérémie F. Cohen,^{1,6}
Martin Chalumeau,^{1,2,6}
Valentine Brousse,^{1,2} and Mariane
de Montalembert^{1,2}

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⁵Établissement Français du Sang (EFS), Necker Hospital for Sick Children, and ⁶Obstetrical, Perinatal and Paediatric Epidemiology Research Team (EPOPÉ), Inserm UMR1153, Paris, France

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Summary

Patients with sickle cell disease (SCD) show a high prevalence of red blood cell (RBC) alloimmunization, but few studies have focused on children. We aimed to study the prevalence and risk factors of RBC alloimmunization in SCD children. We retrospectively analysed the medical and transfusion files for 245 SCD children hospitalized in our centre in 2014 and included 175 patients who had received at least one RBC unit in their lifetime. The main clinical and immuno-haematological characteristics of alloimmunized and non-alloimmunized patients were compared. The prevalence of alloimmunization was 13.7% [95% confidence interval (CI) (8.6–18.6)], and 7.4% [95% CI (3.5–11.3)] after excluding the probable irregular natural antibodies (anti-M, anti-Le^a, anti-Le^b, anti-Le^x). Main risk factors for alloimmunization were increased number of RBC units received (median of 65 vs. 10 units per patient; $P = 0.01$) and the presence of one or more red cell autoantibodies (46.2% vs. 4.7%; $P < 0.0001$). The alloimmunization rate was higher for episodically transfused than chronically transfused patients (1.43 vs. 0.24/100 units received; $P < 0.001$). The presence of red cell autoantibodies appears to be a major risk factor for alloimmunization in SCD children and could justify specific transfusion guidelines.

Keywords: sickle cell disease, children, blood transfusion, red blood cell alloimmunization, immuno-haematology.

Transfusion of blood components

- Transfusion of blood components
 - Transfusion of packed red blood cells (PRBC)
 - Description of component
 - Pretransfusion Compatibility testing
 - **Transfusion of fresh frozen plasma (FFP)**
 - **Description of component**
 - **Pretransfusion Compatibility testing**
 - Transfusion of platelets
 - Description of component
 - Pretransfusion Compatibility testing
- Hemovigilance

Transfusion of fresh frozen plasma (FFP)

Description of component

- ❑ Prepared from whole blood or from plasma collected by apheresis
- ❑ Contains normal plasma levels of stable coagulation factors, albumin and immunoglobulins
- ❑ ≥ 70 IU Factor VIIIc level per 100ml and at least similar amounts of the other labile coagulation factors and naturally occurring inhibitors
- ❑ Conservation at -30°C or lower
- ❑ Administration through a 170-200 μm filter



Transfusion of fresh frozen plasma (FFP)

- ❑ Pathogen reduction treatment
 - ❑ Active on known viruses, bacteria, protozoa and contaminating leukocytes but also on unknown transfusion-transmissible agents, they are not active on prions
 - ❑ Current methods: solvent-detergent, methylene blue, amotosalen, and riboflavin
- ❑ **Pretransfusion Compatibility testing**
 - ❑ Patient's ABO blood group determined on two different blood samples collected at different times

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Transfusion of platelet concentrates (PCs)

Description of component

- PCs
 - Apheresis platelets (prepared by apheresis from single donors)
 - Recovered platelets (prepared by separation and pooling of units of platelets from whole blood of multiple donors)
- No difference in haemostatic effect; platelet survival and post-transfusion increment for both preparations
- Storage: max 5 days after collection at $22\pm 2^{\circ}\text{C}$
- Specific preservative solutions bags permitting O_2 and CO_2 so that the pH during storage stays continuously between 6.4 and 7.4



Transfusion of platelet concentrates (PCs)



- Leukodepleted: decreased incidence of alloantibody mediated refractoriness to PC transfusion, CMV transmission and FNHTRs
- Pathogen reduction treatment
 - Maximum storage time could be extended to 7 days with equivalent safety and clinical efficacy (*Lozano et al, 2011; Schlenke et al, 2011*)
 - But belgian law 6/2/2018: 5 days

- **Pretransfusion Compatibility testing:**

- Patient's ABO/ RhD blood groups
- Contamination of PCs by red cells
 - prevention of RhD immunisation by the use of RhD-immune globulin if RhD+ PC are transfused to RhD- patients of child-bearing age or younger

Platelet express on their surface:

- ABO antigens
- Class I HLA antigens
- HPA antigens

Transfusion of platelet concentrates (PCs)

Response to platelet transfusion

- **Platelet recovery or CCI**
- Absolute Increment =
(Post-transfusion platelet count) – (pre-transfusion platelet count)
- Corrected Count Increment (CCI) =
Absolute Increment X Body surface area / number of platelets transfused (10^{11})
CCI : 10-60 minutes after transfusion (Mc Farland, AABB, 2008)

Transfusion of PCs: Response to platelet transfusion

❑ Transfusion platelet refractoriness

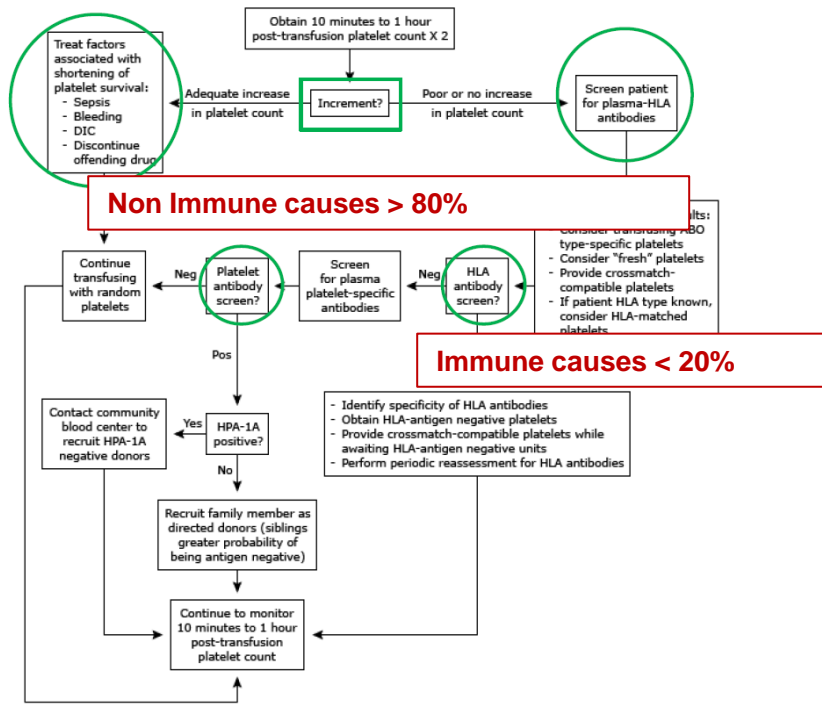
- ✓ Only when at least two ABO-compatible transfusions, stored for <72 hours result in poor increments
- ✓ Defined in a clinically stable patient when two sequential platelet transfusions lead to 1 hour post-transfusion CCIs of less than 5000 platelets x m2 per μ l

- ✓ If \downarrow 1h and 24h: Immune cause (antibodies against HLA class I and/or HPA antigens)
- ✓ If N 1h and \downarrow 24h: Non immune cause (sepsis, splenomegaly, DIC, GvHD, bleeding, medications,...)
- ✓ « *Non immune causes are the most likely and the first that should be explored in the diagnosis of platelet refractoriness* »

Management of the platelet refractory patient. S.K.Forest, E.A.Hod.

Transfusion of platelet concentrates (PCs)

Diagnosis and management of platelet refractoriness



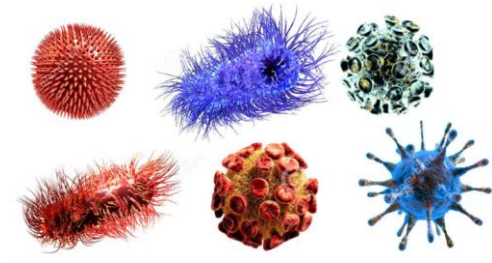
Hemovigilance

- Hemovigilance = Hema (blood in Greek) + Vigilans (watchful in Latin)
- Hemovigilance system improve the safety of blood transfusion by applying a set of surveillance procedures covering whole transfusion chain from the collection of blood and its components to the follow up of its recipients, intended to collect and access information on unexpected or undesirable effects resulting from the therapeutic use of labile blood products, and to prevent their occurrence and recurrence.

Early 1980s...

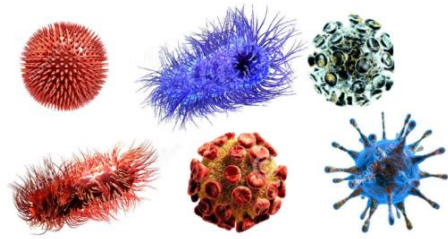


Mistransfusions



Infectious diseases

Today...

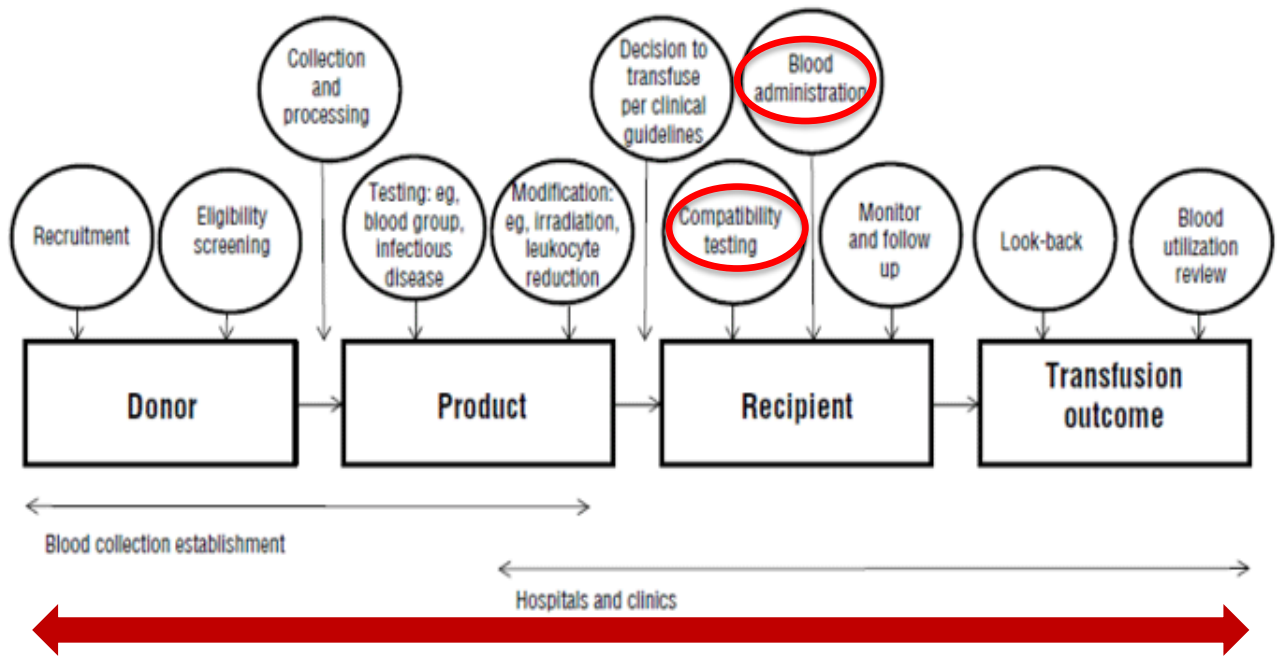


Infectious diseases



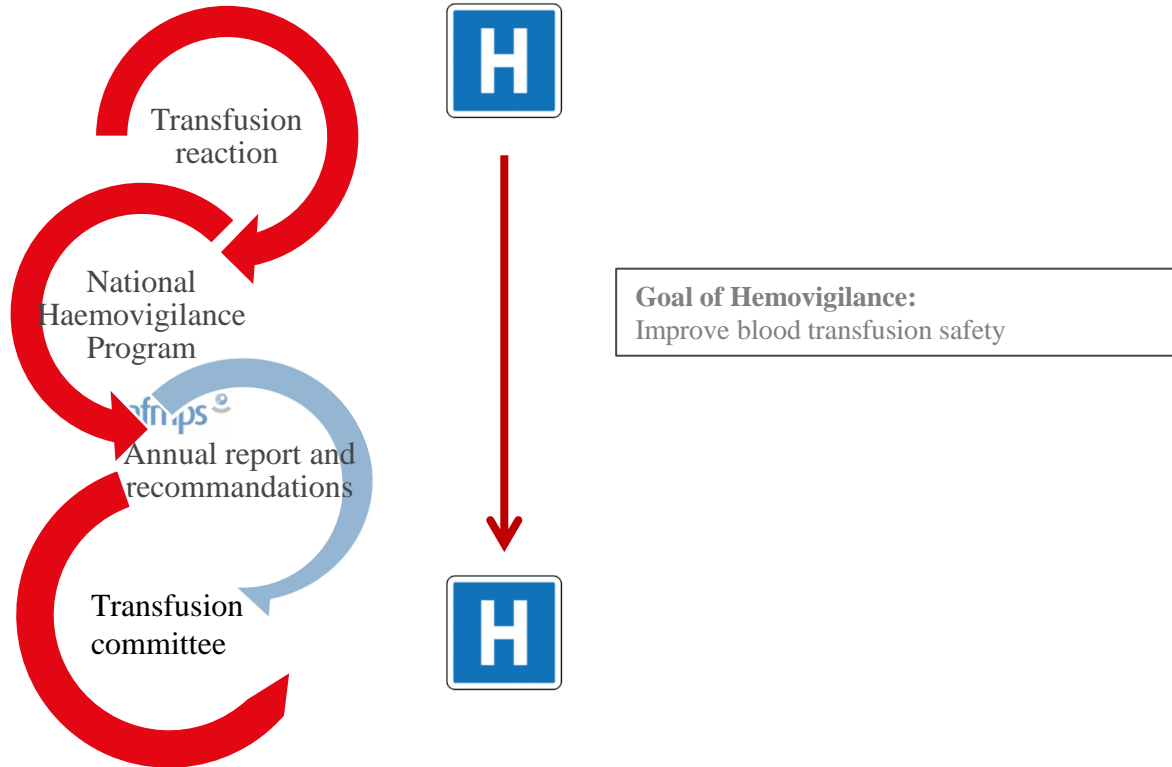
Mistransfusions

Transfusion process

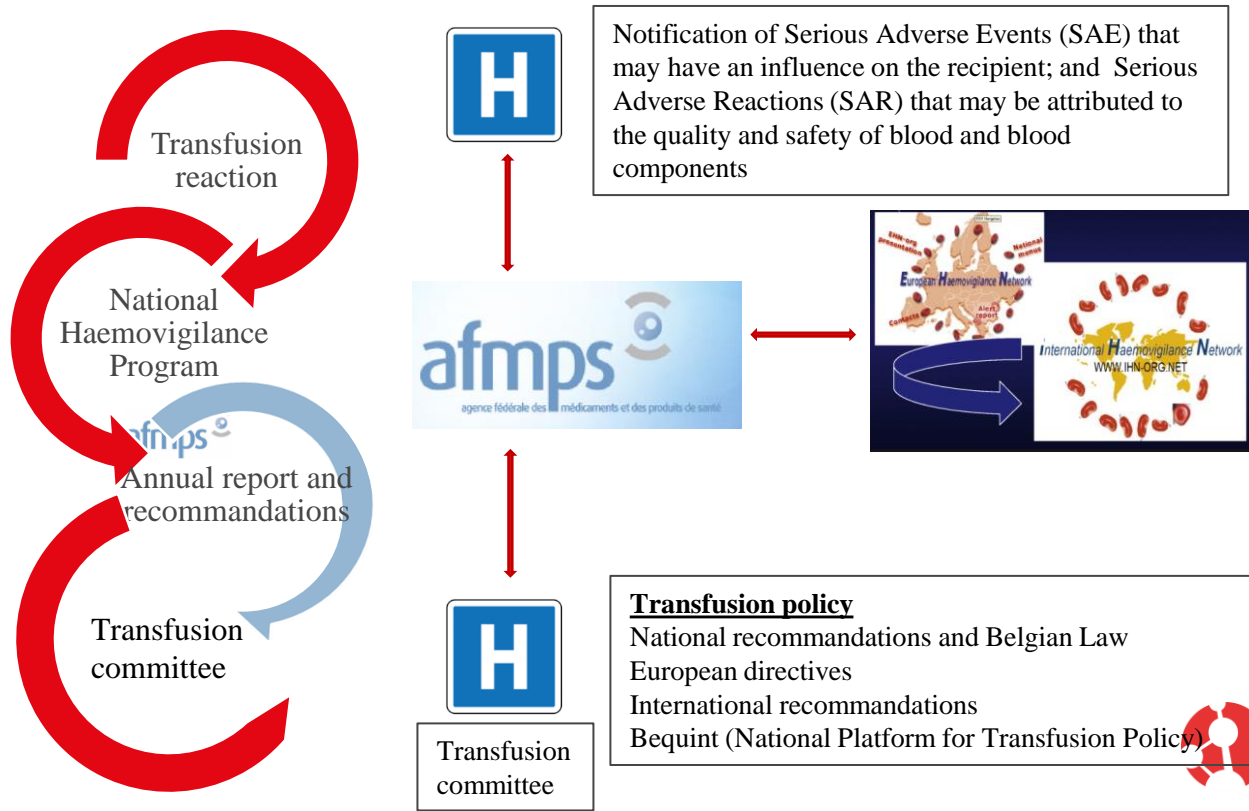


Hemovigilance surveillance programme

Hemovigilance (recipient)



Hemovigilance (recipient)



Hemovigilance: notification form

 Agence Fédérale des Médicaments et Produits de Santé		Département Vigilance Centre d'hémovigilance	
FORMULAIRE DE NOTIFICATION/CONFIRMATION D'UNE RÉACTION TRANSFUSIONNELLE INDESIRABLE GRAVE OU D'UN INCIDENT INDESIRABLE GRAVE EN RAPPORT AVEC UN COMPOSANT SANGUIN			
Code Hôpital Année	Nom Hôpital Numéro d'ordre	<input type="checkbox"/> Notification	<input type="checkbox"/> Confirmation
A. NOTIFICATION			
1. Patient		1.1. date de naissance:	
2. Composant sanguin transfusé		1.2. sexe : M <input type="checkbox"/> F <input type="checkbox"/>	
<input type="checkbox"/> Concentré érythrocytaire (CE) ; numéro d'unité:		<input type="checkbox"/> Allotérique <input type="checkbox"/> Autologue	
<input type="checkbox"/> Concentré plaquettaire (CP) ; numéro d'unité:		<input type="checkbox"/> CP d'aphérèse (donneur urique) <input type="checkbox"/> CP standard	
<input type="checkbox"/> Plasma frais congelé, vira-inactivé (PFCV) ; numéro d'unité:			
<input type="checkbox"/> Autre:		<input type="checkbox"/> plusieurs/différents composants; numéro(s) d'unité:	
3. Réaction transfusionnelle			
3.1. Date et heure : Date et heure du début de la transfusion (en cas de réaction transfusionnelle): _____ à _____ (hr:min)			
Date et heure de la réaction ou de l'incident : _____ à _____ (hr:min)			
3.2. Lieu de la transfusion : <input type="checkbox"/> Bloc opératoire <input type="checkbox"/> Soins intensifs <input type="checkbox"/> Chirurgie <input type="checkbox"/> Hémato-oncologie			
<input type="checkbox"/> Médecine interne <input type="checkbox"/> Pédiatrie <input type="checkbox"/> Hôpital de jour <input type="checkbox"/> Autre : _____			
3.3. Symptômes			
<input type="checkbox"/> malaise	<input type="checkbox"/> tachycardie	<input type="checkbox"/> oligurie, anurie	
<input type="checkbox"/> insécon	<input type="checkbox"/> hypertension	<input type="checkbox"/> hypotension	
<input type="checkbox"/> fièvre	<input type="checkbox"/> arythmie cardiaque	<input type="checkbox"/> choc	
<input type="checkbox"/> démangeaison	<input type="checkbox"/> ictericité	<input type="checkbox"/> perte de connaissance	
<input type="checkbox"/> urticaire	<input type="checkbox"/> douleur thoracique/abdominale	<input type="checkbox"/> hémorragie diffuse	
<input type="checkbox"/> rougeur	<input type="checkbox"/> roudeurs/rouissements	<input type="checkbox"/> autre:	
<input type="checkbox"/> éruption	<input type="checkbox"/> dyspnée	<input type="checkbox"/> autre:	
<input type="checkbox"/> ictere	<input type="checkbox"/> hémoglobinurie	<input type="checkbox"/> autre:	
4. Diagnostic ou syndrome/incident			
<input type="checkbox"/> 4.1. Réaction hémolyse non hémolytique (RHNH)	<input type="checkbox"/> 4.11. infection virale transmise par transfusion :		
<input type="checkbox"/> 4.2. Hémolyse immunologique due à une incompatibilité ABO	<input type="checkbox"/> VIH1,2 <input type="checkbox"/> VIH3		
<input type="checkbox"/> 4.3. Hémolyse immunologique due à un autre allo-anticorps	<input type="checkbox"/> VHC <input type="checkbox"/> CMV <input type="checkbox"/> Autre		
<input type="checkbox"/> 4.4. Hémolyse non immunologique	<input type="checkbox"/> 4.12. Infection parasitaire transmise par transfusion:		
<input type="checkbox"/> 4.5. Purpura post-transfusionnel	<input type="checkbox"/> Malaria <input type="checkbox"/> Autre:		
<input type="checkbox"/> 4.6. Réaction allergique grave	<input type="checkbox"/> 4.13. Dœdème pulmonaire aigu (détaillez le caractère, surcharge volumétrique)		
<input type="checkbox"/> 4.7. Réaction anaphylactique	<input type="checkbox"/> 4.14. Autres réactions indésirables graves:		
<input type="checkbox"/> 4.8. Lésion pulmonaire aiguë post-transfusionnelle (TRALI)	<input type="checkbox"/> 4.15. Administration erronée d'un composant sanguin (veuillez aussi remplir la section 10)		
<input type="checkbox"/> 4.9. Maladie du greffon contre l'hôte	<input type="checkbox"/> 4.16. Near miss (veuillez aussi remplir la section 10)		
<input type="checkbox"/> 4.10. Infection bactérienne post-transfusionnelle (bactériémie, sepsis, choc endotoxique).			
<input type="checkbox"/> Microorganismes(s): _____			
Information complémentaire:			
5. Gravité		6. Imputabilité de l'incident transfusionnel	
<input type="checkbox"/> 0: absence de manifestation clinique	<input type="checkbox"/> 0: exclu	<input type="checkbox"/> 0: improbable	
<input type="checkbox"/> 1: absence de menaces vitales, y compris à long terme	<input type="checkbox"/> 1: non évaluable	<input type="checkbox"/> 1: non évaluable	
<input type="checkbox"/> 2: menace vitale à long terme	<input type="checkbox"/> 2: probable	<input type="checkbox"/> 2: probable	
<input type="checkbox"/> 3: menace vitale immédiate	<input type="checkbox"/> 3: certain, prouvé	<input type="checkbox"/> 3: certain, prouvé	
<input type="checkbox"/> 4: décès			
7. Enquête transfusionnelle		8. Évolution du patient après l'incident (si connue)	
<input type="checkbox"/> en cours	<input type="checkbox"/> rétablissement complet	<input type="checkbox"/> rétablissement complet	
<input type="checkbox"/> terminée	<input type="checkbox"/> séquelles mineures, loquaces:	<input type="checkbox"/> séquelles mineures, loquaces:	
<input type="checkbox"/> non réalisée	<input type="checkbox"/> séquelles graves, loquaces:	<input type="checkbox"/> séquelles graves, loquaces:	
<input type="checkbox"/> non réalisable	<input type="checkbox"/> décès	<input type="checkbox"/> décès	
9. Personne de contact de l'hôpital pour l'hémovigilance			
Validé/signé par la personne de contact (cliquez le champ suivant):			
Nom prénom: _____		<input type="checkbox"/> La réaction peut être imputable à un problème de qualité ou à la sécurité du composant sanguin.	
Date JJ/MM/AAAA: _____		<input type="checkbox"/> La réaction est/à aussi (été) notifiée au centre de transfusion sanguine	

Hemovigilance: AFMPS annual reports



The screenshot shows the AFMPS website interface. At the top, there is a navigation bar with links for 'A propos de l'AFMPS', 'Travailler à l'AFMPS', 'Publications', 'Presse', 'Contact', 'Plaintes', and 'Portail web'. Below this is the AFMPS logo and the tagline 'agence fédérale des médicaments et des produits de santé'. A search bar is also present. The main navigation menu includes 'Usage humain' (highlighted), 'Usage vétérinaire', 'Information pour le public', and 'Information pour le professionnel'. A breadcrumb trail reads: 'Accueil > Usage humain > Produits de santé > Sang et produits sanguins > Hémovigilance > Rapports annuels'. The 'Rapports annuels' section lists several reports, including one on temporary exclusion criteria for blood donors and several annual hemovigilance reports from 2016 to 2019.

Rapports annuels

Rapport critères d'exclusion temporaire don de sang

- [Rapport de la concertation annuelle du 6 décembre 2022 sur les critères d'exclusion temporaire, et les périodes d'exclusion connexes, pour les donneurs concernant le comportement sexuel.](#)
- [Rapport de la troisième concertation annuelle du 15 décembre 2021 sur les critères d'exclusion temporaire, et les périodes d'exclusion connexes, pour les donneurs concernant le comportement sexuel.](#)
- [Rapport de la deuxième concertation annuelle du 8 décembre 2020 sur les critères d'exclusion temporaire, et les périodes d'exclusion connexes, pour les donneurs concernant le comportement sexuel.](#)
- [Rapport de la première concertation annuelle du 10 décembre 2019 sur les critères d'exclusion temporaire, et les périodes d'exclusion connexes, pour les donneurs concernant le comportement sexuel.](#)

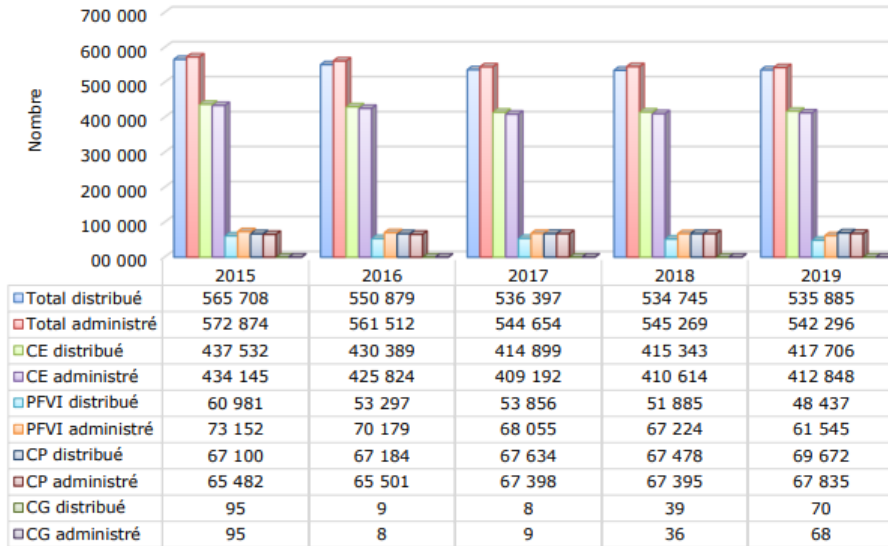
Rapport annuel Hémovigilance

- [Rapport annuel 2019](#)
- [Rapport annuel 2018](#) (corrections 26/11/2021: pages 23 et 28)
- [Rapport annuel 2017](#)
- [Rapport annuel 2016](#)

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4.2. Composants sanguins distribués et administrés

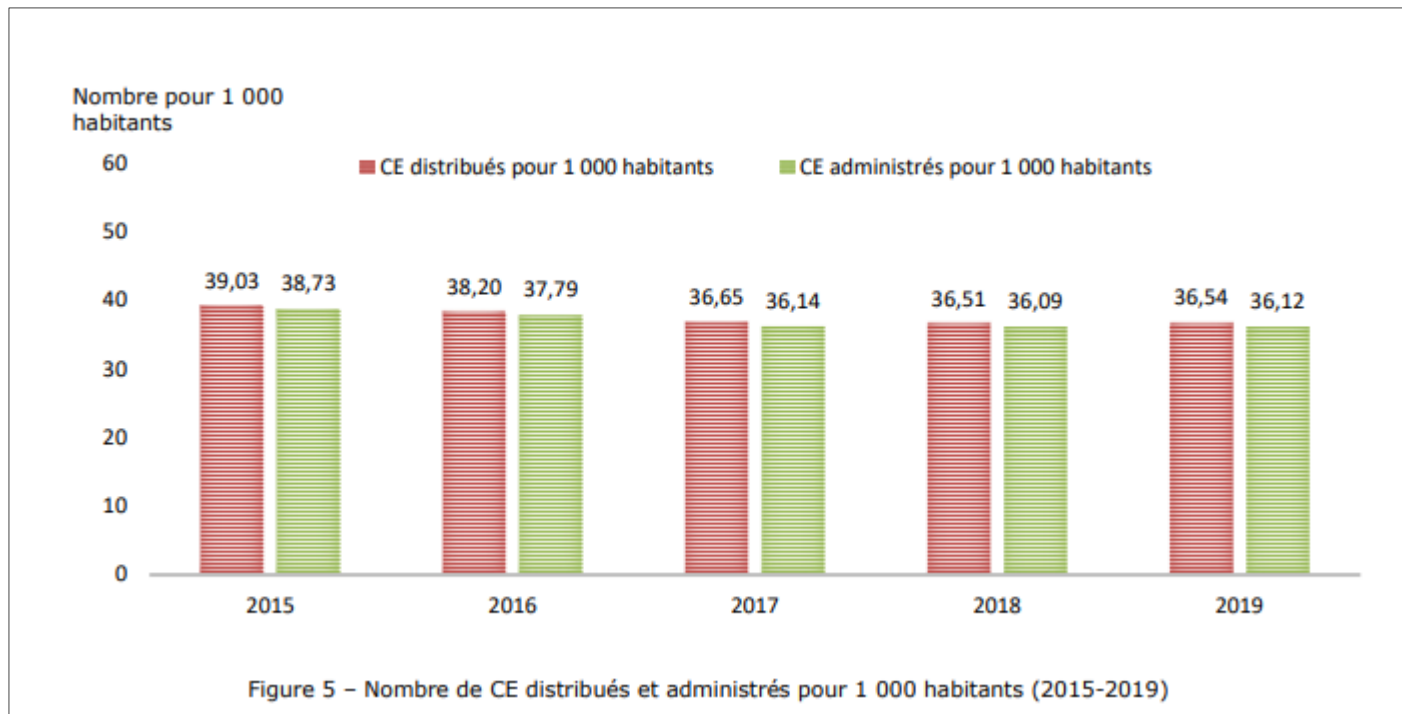
La figure 4 montre l'évolution du nombre de composants sanguins distribués et administrés sur la période 2015-2019. La distribution et l'administration globales des composants sanguins en 2019 sont similaires à l'année précédente.



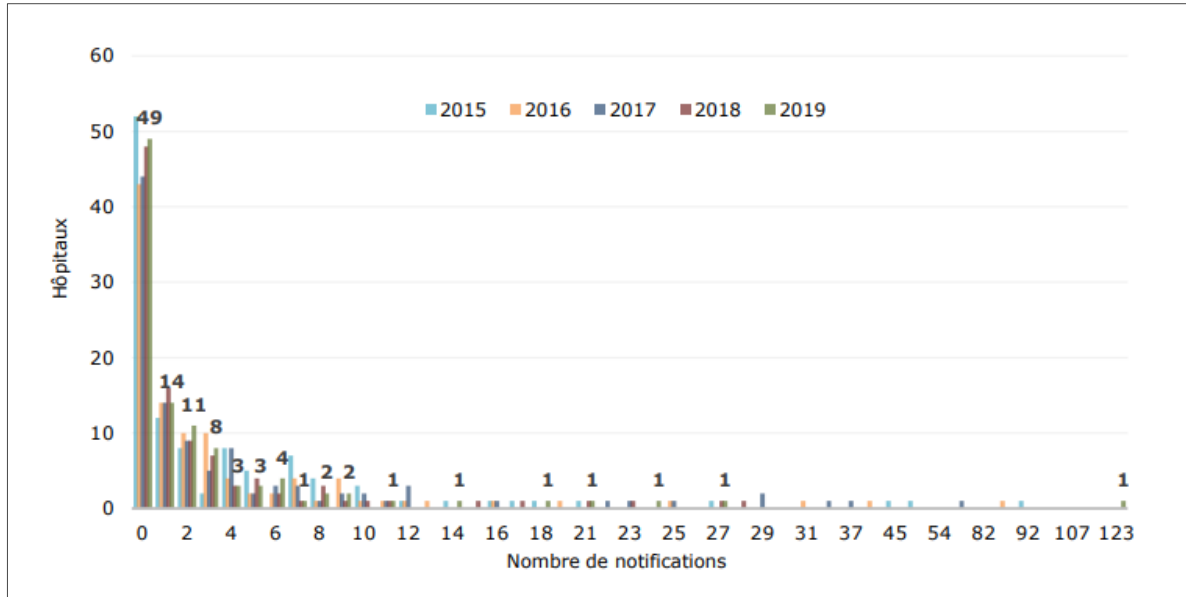
(PFVI = plasma frais viro-inactivé, CG = concentré granulocytaire, CP = concentré plaquettaire)

Figure 4 – Nombre de composants sanguins distribués et administrés (2015-2019)

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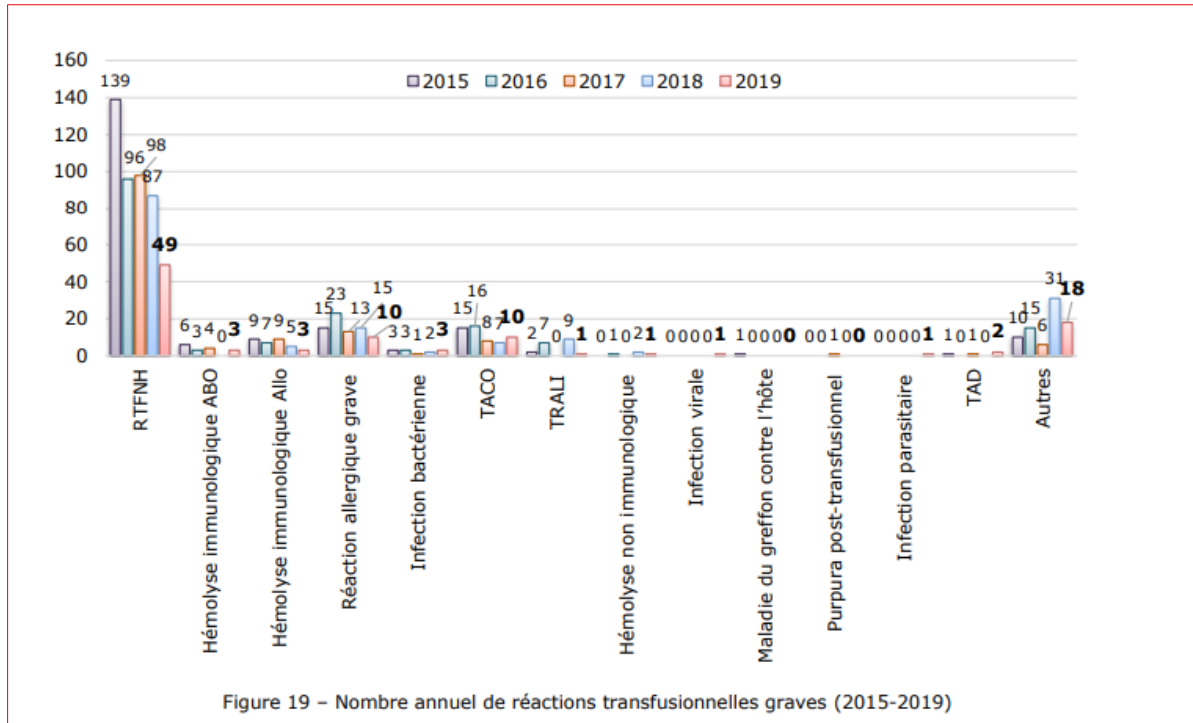


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En 2019, 55 hôpitaux ont notifié au moins 1 incident ou réaction indésirable grave et 49 hôpitaux n'ont notifié aucun incident ou réaction. Le nombre de notifications par hôpital varie de 0 à 123 et le nombre total de notifications par hôpital pour 1 000 composants sanguins administrés varie de 0 à 5,05 (médiane : 0,15).

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Thank you for your attention

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